

Sequencing

Modified by Zhaoyang Liu, April 2016

1. PCR reaction

Aliquot 50ul Big Dye into 1.5ml Eppendorf tube and store at -20C.

Set up the PCR reaction with Big Dye as follows:

Sequencing primer: 2ul (2uM)
5x Big Bye buffer: 2ul
DNA template: ~200-400ng
Big Dye: 1ul
Add H2O to 10ul

PCR program: 96 C 1min
96 C 10s
50 C 5s 25 cycles
60 C 10s
12 C ∞

2. PCR product purification

Aliquot 500ul HiDi buffer into 1.5ml Eppendorf tube and store at -20C.

- Transfer the 10ul of PCR product into a 1.5ml Eppendorf tube.
- Add 1ul 125mM EDTA and 2 volume (20ul) 100% EtOH.
- Centrifuge at >1200g for 15min at room temperature. (Note: 4C works better)
- Carefully remove supernatant. Add 100ul 70% EtOH. (Note: fresh made 70% EtOH works better)
- Centrifuge at >1200g for 5min at room temperature. (Note: 4C works better)
- Carefully remove supernatant. Air dry in dark for at least 20min. (Note: tube must be completely dry!)
- (Note: Samples can be covered with aluminum foil and store at 4C if not use immediately.)
- Add 10ul HiDi buffer and vortex.
- Transfer samples to 96 well PCR plate.
(Note: loading order: A1-H1, A3-H3, A5-H5, A7-H7, A9 -H9, A11-H11; A2-H2, A4-H4.....)

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|----|--|----|--|----|--|----|--|----|--|-----|--|
| A1 | | A3 | | A5 | | A7 | | A9 | | A11 | |
| B1 | | B3 | | B5 | | B7 | | B9 | | B11 | |
| C1 | | C3 | | C5 | | C7 | | C9 | | C11 | |
| D1 | | D3 | | D5 | | D7 | | D9 | | D11 | |
| E1 | | E3 | | E5 | | E7 | | E9 | | E11 | |
| F1 | | F3 | | F5 | | F7 | | F9 | | F11 | |
| G1 | | G3 | | G5 | | G7 | | G9 | | G11 | |
| H1 | | H3 | | H5 | | H7 | | H9 | | H11 | |

3. Sequencing

- Log into the computer with username: HSCADMINSTRATOR, password: Password1
- Turn on 3730 DNA analyzer instrument
- Launch Run3730 Data Collection V2.0 on desktop. (Note: a service console will open. It contains 4 icons that will change from red/yellow to green. Don't close the console window when the software is running.) Login Name: 3730user; Password: 3730users
- In the GA Instruments tab click "plate manager" and fill in as follows:
 - a. click "new" at the bottom left of the window
 - b. enter plate ID (barcode) and plate name. (Note: No space. These two ID can be the same.)
 - c. choose application. For sequencing, choose "sequencing analysis".

- d. choose plate type: 96 well plate
 - e. choose plate sealing: septa
 - f. enter names of owner and operator
 - g. click OK. A sample sheet will show up. Fill in sample names, as well as:
 - h. Result Group: SeqScape
 - i. instrument protocol: BigDye 3.1
 - j. Analysis protocol: Bigdye 3.1
- (Note: for column h to j, choose the whole column and click Ctrl+D. The whole column will be filled)

- Click Run Scheduler
- Search your plate ID to find your plate.
- Once finding your plate, click add samples, done.
- Put your plate into the stackers.
- Click the Green arrow below the file tab to start.

4. Sequencing data analysis

- Click Sequencing Analysis software on the desk top. Username: finnelli, Password: finnelli01
- Click file, add samples. Find your samples in E/3730_Runs/Seq_Result_Group.
- Add all samples. Click the green arrow to start analysis.
- A new data file will be generated after the analysis. Copy all files into your USB drive.
- Open the trace file with the software SeqScanner (Applied Biosystem)